

## Effect of Erythropoietin on Some aspects of Carbohydrate and Lipid metabolism in Obese and diabetic rats

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### Keywords

- Erythropoietin
- Diabetes
- Obesity
- HOMA-I/R
- HOMA S %

### Abstract

**Objectives:** To study effect of erythropoietin (EPO) on some aspects of carbohydrate and lipid metabolism in obese and diabetic rats. **Material and methods.** This study was carried on (50) albino female rats; divided into 5 groups, each consisted of 10 rats: Control, obese, diabetic, erythropoietin treated obese group (after induction of obesity it was injected by erythropoietin 600u/kg I.P. once daily every other day for 5 weeks) and erythropoietin treated diabetic group (after induction of diabetes it was treated as previous group). At the end of experimental period serum samples were collected for estimation of: fasting serum glucose, serum insulin, insulin resistance (HOMA IR), insulin sensitivity (HOMA S%), serum LDL, serum HDL, serum triglyceride (TG), serum cholesterol, glycosylated Hb% and measurement of body mass Index (BMI) **Results:** Erythropoietin treated obese group showed significant decrease in fasting glucose, insulin, HOMA IR, serum LDL, TG, cholesterol and BMI. While, serum HDL and HOMA S% were significantly increased when these results were compared to obese group. Erythropoietin treated diabetic group produced insignificant change in all studied parameters with exception of significant decrease in fasting glucose, TG and glycosylated Hb% when these results compared to diabetic group. **Conclusion:** EPO can be considered as adjunct anti-diabetic/obesity drug to reduce blood glucose, hypolipidemic effect and attenuate weight gain.

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## INTRODUCTION

In the last few decades, the prevalence of obesity and diabetes has been increasing rapidly (1). There is strong association between obesity, cardiovascular disease, type 2 diabetes and other chronic diseases, this trend suggests a substantial increase in obesity-related morbidity and mortality for the future (2), as regarding diabetes mellitus its effect include long term damage, dysfunction and failure of various organs (3). The increased risk of several co-morbidities associated with obesity and diabetes mellitus increase the need for developing a new applicable treatment.

Erythropoietin (EPO) is a glycoprotein predominantly produced in the kidney and important for production of red blood cell also it acts on several non-erythropoietic tissues. The presence of erythropoietin receptors in cells other than erythroid progenitors suggests that erythropoietin has other biological function in addition to erythropoiesis (4). It has been proved that EPO signaling has a role in regulation of body weight, blood glucose and fat mass (5) in patient with chronic renal disease. EPO administration has shown significant improvement in several metabolic parameters including fasting glucose level and insulin sensitivity (6). Also the findings that pancreatic  $\beta$ -cells harbor functional EPO-Rs and that EPO acts directly on them raise the possibility that EPO treatment may also affect insulin secretion by the pancreatic cells(7).

Previous studies clearly documented the effect of EPO on lipid profile in patient with end stage renal disease by decreasing serum levels of triglycerides, total cholesterol and LDL-

cholesterol(8). Therefore, the aim of the present study was to analyze the effect of EPO on some aspects of carbohydrate and lipid metabolism in obese and diabetic rats.

## MATERIALS AND METHODS

### Animals

The present work was carried on 50 female albino rats ranging in weight between 160-180g, 12-16 weeks old. The rats were housed in isolated animal cages, in a standard animal laboratory room and had free access to water and food all over the period of the work, and kept at room temperature.

The animals were divided into five groups (10 rats in each group):

**Group 1** (control group): The animals of this group which were fed standard ad libitum (free access of animal to food and water) commercial chow with tap water injected i.p. by 1ml saline daily for 5 weeks.

**Group 2** (Obese group): Obesity induced in these rats by feeding a high fat diet composed of 70% fat, 20% carbohydrates and 10% protein. It consists of cooked cow fat, casein, bread and green vegetables for 4 weeks (9). At the end of feeding period their body weights ranged between 220-260g. Rats in this group were injected by 1 ml saline i.p. daily for 5 weeks after induction of obesity.

**Group 3** (Diabetic group): Diabetes induced by multiple low doses of streptozotocin injected i.p. in a dose of 40mg/kg for 5 consecutive days, after a period of two weeks blood sample analysis of rats showed marked hyperglycemia with blood glucose level up to 200 mg/dl (10). Rats in this

group were injected by 1 ml saline i.p. daily for 5 weeks after induction of diabetes.

**Group 4:** (Erythropoietin treated obese group) rats were injected i.p. with erythropoietin in a dose of 600u/kg once daily every other day for 5 weeks after induction of obesity (11).

**Group 5:** Erythropoietin treated diabetic group: rats were injected i.p. with erythropoietin two weeks after the start of streptozotocin injection in a dose of 50µg/kg (600u/kg) once daily every other day for 5 weeks (11). Normal and high fat diet constituents were purchased from El-Gomhoria Company, Cairo, Egypt. High fat diet was preserved at 4°C until used All protocols were approved by Tanta Faculty of medicine ethical Committee e Diets of all groups are equal in amount but different only in there constituent

#### **Biochemical assay:**

At the end of the experimental period, the animals were fasted overnight, rats were anesthetized by i.p. injection of pentobarbital sodium (50mg/Kg body weight) (12). Then, body weight and body length (nose –anus length) (13) were measured for all groups, blood sample were collected by decapitation of rats and centrifuged at 3000 rpm for 10 minutes and the separated serum was then transferred into clean storage tubes for estimation of the following parameters except for estimation of glycosylated hemoglobin% and whole blood sample used for determination of fasting glucose level according to method of Tietz (14). Serum insulin level according to method of Kao et al., (15) where intra-assay and inter-assay coefficients of variation<10% and ELISA reader capable of reading absorbance at 450nm. HOMA IR ( $\text{HOMA-I/R} = \text{fasting insulin } (\mu\text{IU/ml}) \times \text{fasting}$

$\text{glucose (mg/dl)}/405)$  and  $\text{HOMA S } \%(1/\log \text{insulin } (\mu\text{IU/ml}) + \log \text{glucose (mg/dl)})$  was measured using homeostasis model of assessment (HOMA) analysis (16). Serum LDL cholesterol was measured according to method of Assmann et al, (17). Serum HDL cholesterol was measured according to method of Grove (18). Serum triglycerides level was measured by GPO enzymatic method McGowan et al., (19). Serum total cholesterol level was measured by BioMed-cholesterol-LS kits according to Tietz, (20). Glycosylated hemoglobin%: was measured according to method of Bissé and Abraham (21). Body mass index (BMI) was measured using the formula of Novelli et al, (13).The kits and biochemical used in the study obtained from Sigma chemical Co.

#### **Statistical analysis:**

The data were expressed as the mean  $\pm$  standard deviation. Data from our study were analyzed using the unpaired student's t-test to assess significant difference between two groups. P-values <0.05 were considered statistically significant. All the analyses were performed using Graph Pad InStat, 32 bit for win 95/NT (Version 3.05).

## **RESULTS**

Our results are shown in tables 1 and 2 and figures 1-4. In the obese group, fasting glucose is insignificantly changed when compared to control group, while serum insulin, HOMA IR, serum LDL, serum triglyceride, serum cholesterol and BMI showed significant increase when compared with the control group. On the other hand HOMA S% and serum HDL in the obese group showed

significant decrease when compared with control group (table 1 and figures 1 and 2).

In the erythropoietin treated obese group, fasting glucose, serum insulin ,HOMA IR,serum LDL, serum TG, cholesterol and BMI (g/cm<sup>2</sup>) showed significant decrease when compared with the

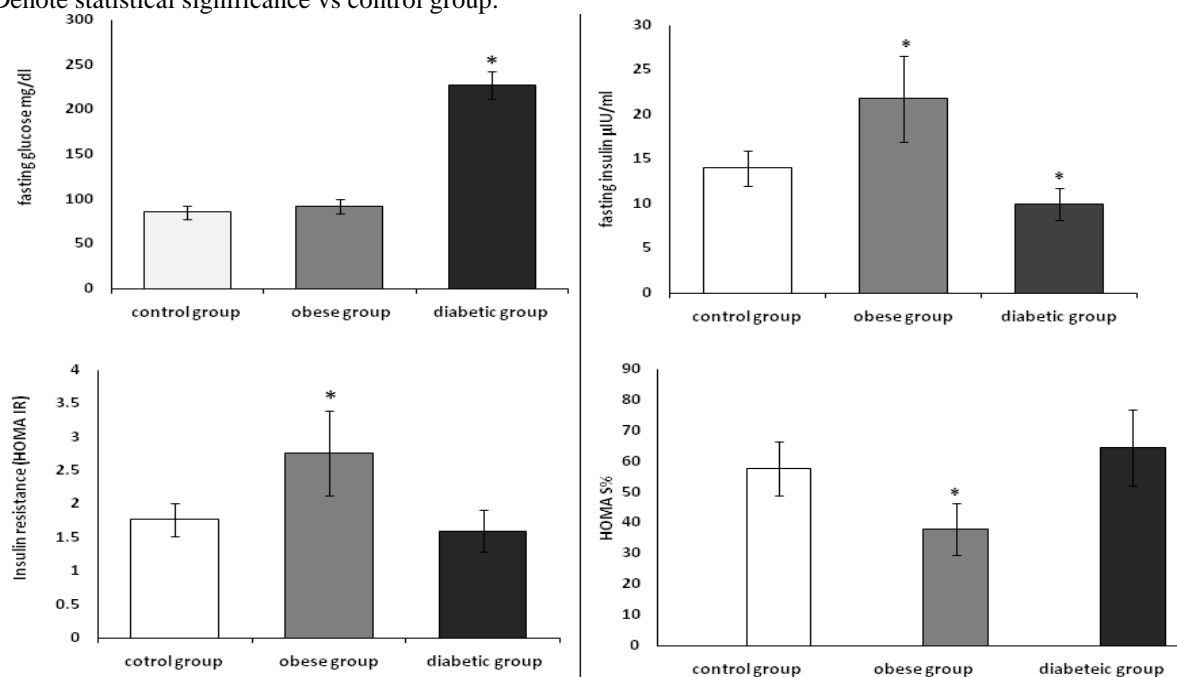
control obese group. A significant increase in HOMA S% serum level of HDL in the erythropoietin treated obese group when compared with the obese group. However, glycosylated Hb results in obese group and in erythropoietin treated obese cannot be statistically analyzed as SD is zero (table 1 and figures 3 and 4).

**Table (1):** Comparison between all parameters studied in control normal, control obese and control diabetic groups

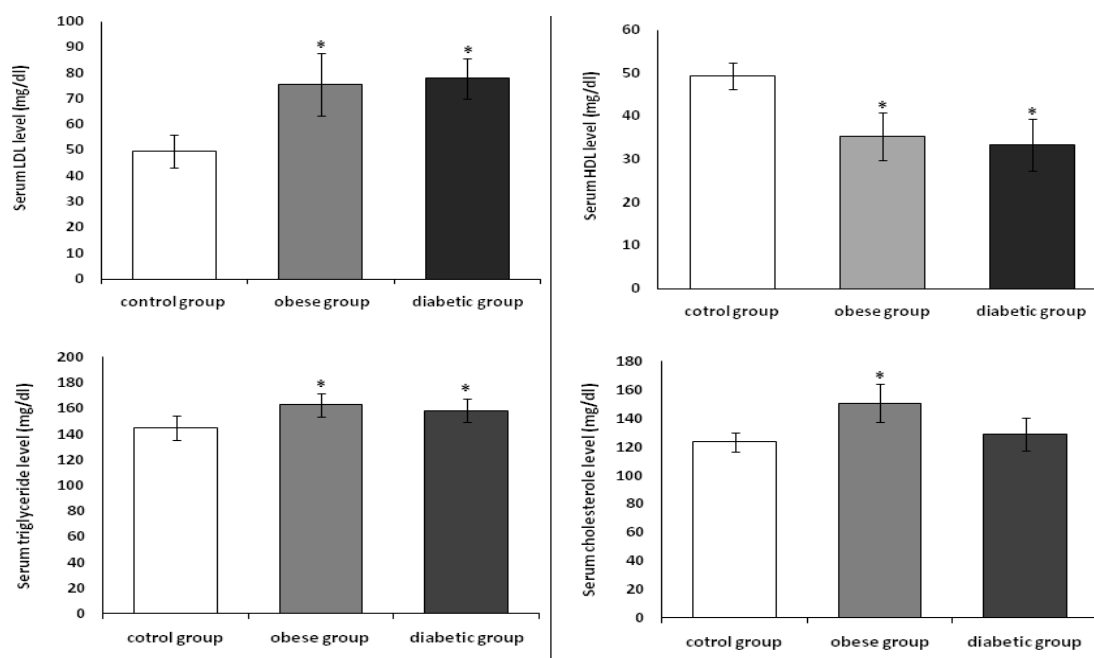
Parameters	Control group (n=10)	Obese group (n=10)	Diabetic group (n=10)	P <sub>1</sub> value	P <sub>2</sub> value
Fasting glucose (mg/dl)	85.48±7.38	91.94±7.70	227.19±15.40*	0.071	0.0001
Serum insulin (µIU/ml)	13.98±2.04	21.76±4.85*	9.98±1.81*	0.0002	0.0001
HOMA IR	1.77±0.25	2.76±0.63*	1.60±0.31	0.0002	0.187
HOMA S%	57.73±8.74	37.83±8.42*	64.37±12.3	0.0001	0.1812
Serum LDL (mg/dl)	49.75±6.48	75.54±12.13*	77.87±7.87*	0.0001	0.0001
Serum HDL (mg/dl)	49.27±3.07	35.29±5.57*	33.38±5.92*	0.0001	0.0001
Serum TG (mg/dl)	144.69±9.19	162.64±8.97*	158.16±9.14*	0.0003	0.0041
Serum cholesterol (mg/dl)	123.34±6.67	150.46±13.41*	128.90±11.40	0.0001	0.1998
BMI (g/cm)	0.56±0.08	0.78±0.07*	0.44±0.05*	0.0001	0.0007

Data are given as mean ± SD. (P<sub>1</sub>) Obese group vs control group & (P<sub>2</sub>) Diabetic group vs control.

\*Denote statistical significance vs control group.



**Fig. (1):** Fasting glucose, fasting insulin, HOMA IR, HOMA S% in control normal, control obese and control diabetic groups. \*Denote statistical significance vs control group.



**Fig. (2):** Serum LDL, HDL, triglycerides and cholesterol levels in control normal, control obese and control diabetic groups. \*Denote statistical significance vs control group.

In diabetic group the fasting glucose, serum LDL, serum triglyceride showed significant increase when compared to the control group. On the other hand serum insulin, serum HDL and BMI in the diabetic group showed significant decrease when compared to control group. However the HOMA IR, HOMA S% and cholesterol in the diabetic group are insignificant when compared to control group, in diabetes, the glycosylated Hb results cannot be statistically analyzed as SD is zero for control group (table 2).

In erythropoietin diabetic group treated group, The fasting glucose, serum level of TG and glycosylated Hb% showed significant decrease

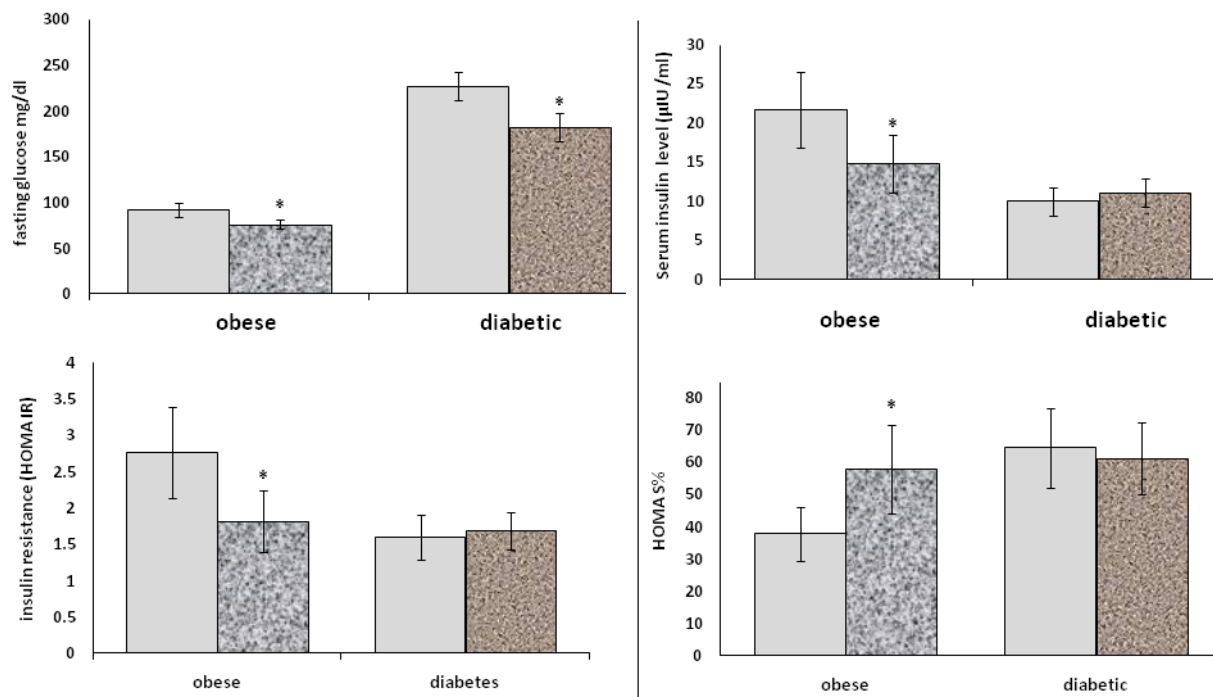
when compared with the diabetic non treated group, serum insulin, HOMA S%, serum LDL level, serum HDL, cholesterol and BMI showed insignificant change when compared to the control diabetic group (table 2).

## DISCUSSION

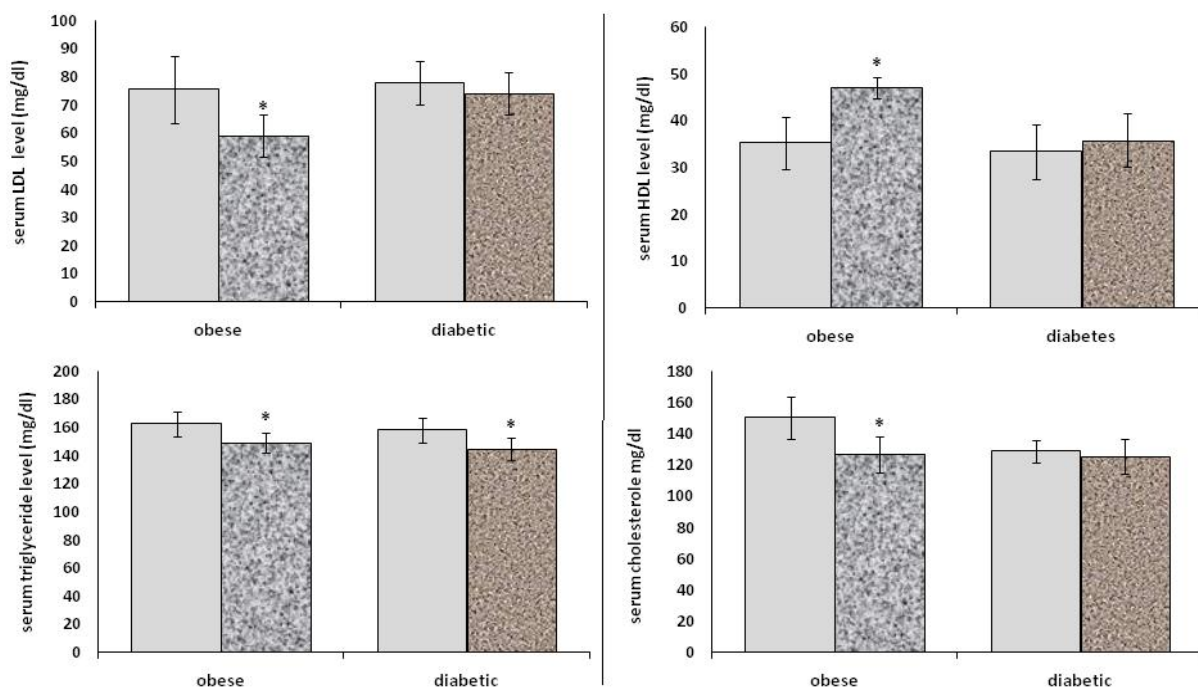
Induction of obesity in rats results in a picture of metabolic syndrome (22). EPO treated obese group showed significant decrease in fasting glucose, serum insulin, HOMA IR, LDL, triglyceride, total cholesterol and BMI. However HDL and HOMA S% were significantly increased, the reduction of the serum glucose level may be due to inhibition of gluconeogenesis by decreasing the expression of phosphoenol-pyruvate carboxykinase (PEPCK) and glucose-6-phosphatase (G6Pase) which are two rate-limiting enzymes for hepatic glucose production(23). Furthermore, EPO administration increases vascular endothelial growth factor(VEGF) expression in the pancreatic islets which mediates angiogenesis in the islets via cell JAK2 pathway (24). JAK2 is able to phosphorylate and activate the downstream antiapoptotic target signals (25). Also, the lowering of glucose level could be due to that EPO induced increase in glucose uptake by increasing

glucose transporter 4 (GLUT4) trafficking toward the plasma membrane in the adipocyte cells

(26).EPO also may lower the fasting glucose by increasing sensitivity to insulin.



**Fig. (3):** Fasting glucose, fasting insulin, HOMA IR, HOMA S% in obese, erythropoietin treated obese, diabetic and erythropoietin treated diabetic groups. \*Denote statistical significance vs control group.



**Fig. (4):** Serum LDL, HDL, triglycerides and cholesterol levels in obese, erythropoietin treated obese, diabetic and erythropoietin treated diabetic groups. \*Denote statistical significance vs control group.

**Table (2):** Comparison between all parameters studied in erythropoietin treated obese and erythropoietin treated diabetic groups

Parameters	Obese group (n=10)	Erythropoietin treated obese group (n=10)	Diabetic group (n=10)	Erythropoietin treated diabetic group (n=10)	P <sub>1</sub> value	P <sub>2</sub> value
Fasting glucose (mg/dl)	91.94±7.70	75.83±4.84*	227.19±15.40	182.21±14.96 <sup>#</sup>	0.0001	0.0001
Serum insulin (µIU/ml)	21.76±4.85	14.91±3.70*	9.98±1.81	11.14± 1.76	0.0023	0.1645
HOMA IR	2.76±0.63	1.82±0.43*	1.60±0.31	1.69±0.26	0.0010	0.4872
HOMA S%	37.83±8.42	57.91±13.64*	64.37±12.31	61.08±11.16	0.0009	0.5391
Serum LDL (mg/dl)	75.54±12.13	59.30±7.47*	77.87±7.87	74.19± 7.59	0.0020	0.3004
Serum HDL (mg/dl)	35.29±5.57	47.12±2.22*	33.38±5.92	35.81±5.67	0.0001	0.3609
Serum TG(mg/dl)	162.64±8.97	149.27±7.16*	158.16±9.14	144.68±7.96 <sup>#</sup>	0.0017	0.0025
Serum cholesterol (mg/dl)	150.46±13.41	126.96±7.05*	128.90±11.40	125.71± 11.25	0.0001	0.5371
BMI (g/cm)	0.78±0.70	0.59±0.05*	0.44±0.05	0.42±0.05	0.0001	0.2751
Glycosylated Hb (%)	----	----	7.4%±0.52	6.4%±0.52 <sup>#</sup>	-----	0.0004

Data are given as mean ± SD. (P<sub>1</sub>) erythropoietin treated obese group vs control obese. (P<sub>2</sub>) Erythropoietin treated diabetic group vs control diabetic. \*Denote statistical significance vs control obese. <sup>#</sup>Denote statistical significance vs control diabetic. It is not possible to analyze Glycosylated Hb (%) in erythropoietin treated obese group vs control obese because SD is zero.

Results of the present work revealed significant decrease in serum insulin and HOMA- IR when EPO is administrated to obese rats which could be due to reducing the transcription factor enhancing binding protein alpha (CEBP/α). This transcription factor play role in insulin sensitivity as it is required during adipogenesis for development of insulin-stimulated glucose uptake (27). EPO therapy significantly decreased plasma cell differentiation antigen 1 (PC-1) activity to the normal value. Elevated levels of PC-1 play a role in the development of insulin resistance in obesity as PC-1 inhibits insulin signaling either at the level of the insulin receptor or downstream at a post

receptor site (28) Also, the mechanism by which EPO affects sensitivity to insulin is that EPO may operate via an increase in NO which is a powerful vasodilator as well as insulin sensitizer (29). Improvement in insulin sensitivity caused by EPO receptor agonist was largely due to enhancement of insulin-stimulated glucose uptake in skeletal muscle and heart by activation of non-oxidative mechanisms of glucose utilization in skeletal muscle and heart (30).

Concerning lipid metabolism it is evident from the present work that EPO when administrated to obese rats causes significant decrease in serum triglyceride, cholesterol, LDL-cholesterol, on the other hand EPO result in significant increase in

HDL-cholesterol. The significant decrease of triglyceride level may be related to an improved response to insulin, since it is known that patients with increased insulin resistance had diminished lipoprotein lipase activity, while the triglyceride production remains normal and thus results in hypertriglyceridemia (6).

Siamopoulos et al., (31) reported that the increase in serum HDL levels could be attributed to the improvement in tissue oxygenation which increases activity of ATP-binding cassette transporter (ABCA1) or other enzymes involved in HDL maturation and lead to the increase in HDL levels. It is known that ABCA1 mediates the efflux of cholesterol and phospholipids to lipid-poor apolipoproteins (apo-A1 and apo-E), which then form HDL (32). Epo treatment improves glucose utilization and reduces insulin resistance, and it is known that high insulin plasma levels stimulate cholesterol synthesis (33). As regard the BMI it is evident from the present work that EPO significantly reduce the BMI in obese group, increased energy consumption or expenditure is a possible mechanism (34).

It was reported that rhEPO not only increased the serum level of leptin, but up-regulated the expression levels of hepatic leptin receptors (Ob-Ra and Ob-Rb). Therefore, it is accepted that EPO might be involved in leptin-mediated weight loss (35). EPO treated diabetic group produced insignificant change in all studied parameters with exception of significant decrease in fasting glucose, serum triglyceride and glycosylated Hb% when these results compared to control diabetic group. As regard to the hypoglycemic effect of

EPO could be due to promotion of  $\beta$ -cell growth and survival it inhibits apoptosis in  $\beta$  cells during diabetes progression (36). Glycosylated Hb % is an indicator for improvement of blood glucose level in diabetic (37), thus the reduction of glycosylated Hb% could be due to the hypoglycemic effect of the EPO in the present work. The significant decrease of triglyceride level may be related to an improved response to insulin (6)

### Conclusion

It can be concluded that EPO can be considered as adjunct anti-diabetic/obesity drug to reduce blood glucose, hypolipidemic effect and attenuate weight gain.

### REFERENCES

1. **Keller U:** From obesity to diabetes. International Journal for Vitamin and Nutrition Research 76: 172–7, 2006.
2. **Maggio AB, Martin XE, Saunders Gasser C, Gal-Duding C, Beghetti M, Farpour-Lambert NJ, et al** Medical and non-medical complications among children and adolescents with excessive body weight BMC Pediatr-14(1):232, 2014
3. **Pokhriyal BN, Thorat K, Dubey R, Limaye D A, Joshi YM, and Kadam VJ,** Urine C-peptide test: A novel non-invasive technique for diagnosis and management of patients with type-1 diabetes IJRPC 220:446-51, 2012
4. **Arcasoy MO:** The non-hematopoietic biological effects of erythropoietin. Br J Haematol 2: 14–31, 2008.



5. **Teng R, Gavrilova O, Suzuki N, Chanturiya T, Schimel D, et al.** Disrupted erythropoietin signalling promotes obesity and alters hypothalamus proopiomelanocortin production. *Nat Commun.* Nov 1; 2:520, 2011.
6. **Khedr E, El-Sharkawy M, Abdulwahab S, Eldin EN, Ali M, et al.** Effect of recombinant human erythropoietin on insulin resistance in hemodialysis patients. *Hemodial Int* 13: 340–6. 2009.
7. **Fenjves ES, Ochoa MS, Cabrera O, Mendez AJ, Kenyon NS, Inverardi L & Ricordi C** Human, nonhuman primate, and rat pancreatic islets express erythropoietin receptors. *Transplantation* 75:1356–60, 2003.
8. **Siamopoulos KC., Gouva C., Katopodis KP., Tzallas C., Nikolopoulos P., Papavasiliou EC et al:** Long-Term Treatment With EPO Increases Serum Levels of High-Density Lipoprotein in Patients With CKD. *American Journal of Kidney Diseases*; 48: 242-9, 2006.
9. **Gong HX., Guo XR., Fei L., Guo M., Liu QQ., Chen RH.,:** Lipolysis and apoptosis of adipocytes induced by neuropeptide YY 5 receptor antisense oligodeoxynucleotides in obese rats. *Acta Pharmacol. Sin.* 2003; 24: 569-75.
10. **Choi D., Schroer SA., Lu SY., Wang L., Wu X., Liu Y., et al** Erythropoietin protects against diabetes through direct effects on pancreatic  $\beta$  cells. *J. Exp. Med.* 207: 2831-42, 2010.
11. **Foskett A., Alnaeeli M., Wang L., Teng R., and Noguchi CT:** The Effects of Erythropoietin Dose Titration during High-Fat Diet-Induced Obesity. *J Biomed Biotechnol*; Volume 2011: Article ID 373781 ,2011
12. **Chen MM., Ashley EA., Deng DX., Tsalenko A., Deng A., and Tabibiazar R.,** :Novel role for the potent endogenous inotrope apelin in human cardiac dysfunction. *Circulation* 108: 1432–9, 2003.
13. **Novelli EL., Diniz YS., Galhardi CM., Ebaid GM., Rodrigues HG., ManiF., et al:** Anthropometrical parameters and markers of obesity in rats. *Lab Anim* 41: 111-9,2007
14. **Tietz NW:** Determination of blood glucose, *Text book of clinical chemistry* WB Saunders. Co. London, Philadelphia; 796, 1986.
15. **Kao P.C., Taylor R.L. and Service F.J.** Proinsulin by Immunochemiluminometric Assay for the Diagnosis of Insulinoma. *Jorunal of Clinical Endocrinology and Metabolism* 78: 1048-51, 1994.
16. **Matthews DR., Hosker JP., Rudenski AS., Naylor BA., Treacher DF., and Turner RC.,** Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 28: 412–9, 1985.
17. **Assmann G., Jabs HU., Kohnert U., Nolte W., and Schriewer H:** LDL-cholesterol determination in blood serum following precipitation of LDL with polyvmylsulfate. *Clin Chim Acta* 140: 77-83,1984.
18. **Grove TH:** Effect of reagent pH on determination of high-density lipoprotein cholesterol by precipitation with sodium phosphotungstate-magnesium. *Clin Chem* 25: 560-4, 1979.
19. **Mcgowan MW., Artiss JD., Strandbergh DR., Zak B:** A peroxidase-coupled method

- for the colorimetric determination of serum triglycerides. *Clin Chem* 29: 538-42, 1983.
20. **Tietz, N.W. ed.**, *Clinical Guide to Laboratory Tests*, 3rd Edition, W.B. Saunders, Co., Philadelphia. 578-580,1995
  21. **Bissé E. and Abraham EC**: New less temperature-sensitive microchromatographic method for the separation and quantitation of glycosylated hemoglobins using a non-cyanide buffer system. *J Chromatog* 344: 81-91, 1985.
  22. **Sutherland LN, Capozzi LC, Turchinsky NJ, Bell RC, Wright DC**: Time course of high-fat diet-induced reductions in adipose tissue mitochondrial proteins: potential mechanisms and the relationship to glucose intolerance. *American Journal of Physiology*. 295:1076–83, 2008.
  23. **Meng R., Zhu D., Bi Y., Yang D., and Wang Y**: Erythropoietin Inhibits Gluconeogenesis and Inflammation in the Liver and Improves Glucose Intolerance in High-Fat Diet-Fed Mice. *PLOS ONE* 8: 1-9, 2013.
  24. **Reinert RB, Cai Q, Hong JY, Plank JL, Aamodt K, Prasad N, et al.** Vascular endothelial growth factor coordinates islet innervation via vascular scaffolding. *Development*. 141(7):1480-91, 2014
  25. **Rusai K., Prokai A., Szebeni B., Fekete A., Treszl A., Vannay A., et al** : Role of serum and glucocorticoid-regulated kinase-1 in the protective effects of erythropoietin during renal ischemia/reperfusion injury. *Biochem Pharmacol* 79: 1173-81, 2010
  26. **Mikolas E., Cseh J., Pap M., SzijartoIA., Laczy., et al** .Effects of erythropoietin on glucose metabolism *Horm Metab Res* .44(4):279-85,2012
  27. **Cha HC., Oak NR., and Mac Dougald OA** :Phosphorylation of CCAAT/Enhancer-binding Protein  $\alpha$  Regulates GLUT4 Expression and Glucose Transport in Adipocytes. *J Biol Chem* 283: 18002-11, 2008.
  28. **Pan Y, Shu JL, Gu HF, Zhou DC, Liu XL, Qiao QY. et al.** Erythropoietin improves insulin resistance via the regulation of its receptor-mediated signaling pathways in 3T3L1 adipocytes.*Molecular and cellular endocrinology*. 367:116–23, 2013
  29. **Marzo F., Lavorgna A., Coluzzi G., Santucci E., Tarantino F., Rio T., et al**: Erythropoietin in heart and vessels: focus on transcription and signalling pathways. *Journal of Thrombosis and Thrombolysis* 26: 183–7, 2008
  30. **Scully MS., Ort TA., James IE., Bugelski PJ., Makropoulos DA., Deutsch HA., et al .** Novel EPO Receptor Agonist Improves Glucose Tolerance via Glucose Uptake in Skeletal Muscle in a Mouse Model of Diabetes. *Exp Diabetes Res*; 2011: 910159, 2011
  31. **Siamopoulos KC., Gouva C., Katopodis KP., Tzallas C., Nikolopoulos P., Papavasiliou EC., et al**: Long-Term Treatment With EPO Increases Serum Levels of High-Density Lipoprotein in Patients With CKD. *American Journal of Kidney Diseases* 48: 242-9, 2006.
  32. **Kruit JK., Kremer PH., Dai L., Tang R., Ruddle P., de Haan W., et al**: Cholesterol efflux via ATP-binding cassette transporter A1 (ABCA1) and cholesterol uptake via the LDL receptor influences cholesterol-induced

impairment of beta cell function in mice. *Diabetologia* 53:1110-19, 2010.

33. **Kes P, Bobić I, Reiner Z, Ratković-Gusić I;** Effect of erythropoietin therapy on serum lipoprotein levels in patients on hemodialysis. *Lijec vjesn* 124(5):146-50, 2002
34. **Katz O., Stuible M., Golishevski N., Lifshitz L., Tremblay ML., Gassmann M., et al:** Erythropoietin treatment leads to reduced blood glucose levels and body mass: insights from murine models. *Journal of Endocrinology* 205: 87–95, 2010.
35. **Qin L., Xiang Y., Song Z., Jing R., Hu C., and, Howard ST:** Erythropoietin as a possible mechanism for the effects of intermittent hypoxia on bodyweight, serum glucose and leptin in mice. *Regulatory Peptides* 165: 168–73, 2010.
36. **Choi D., Schroer SA., Lu SY., Wang L., Wu X., Liu Y., et al:** Erythropoietin protects against diabetes through direct effects on pancreatic  $\beta$  cells. *J. Exp. Med* 207: 2831-42, 2010.
37. **Little RR., Rohlfing CL., and SacksDB.:** Status of haemoglobin A1c measurements and goals for the improvement: from chaos to order for improving diabetes care. *Clin Chem* 57:205-214, 2011